

tions. Determination of the sensitivity of microorganisms to antimicrobial drugs. Version 2024-02. Year of approval (frequency of revision): 2024; MAKMAKH, SSMUJ. Smolensk: 192. <https://www.antibiotic.ru/minzdrav/category/clinical-recommendations/> (In Russ).

8. Belotserkovskiy B.Z., et al. Etiologicheskaya struktura infektsii u patsientov otdeleniya khirurgicheskoi reanimatsii i intensivnoi terapii v postkovidnuiu epokhu [Etiological structure of infections in patients of the surgical intensive care unit in the post-covid era]. Klinicheskaya mikrobiologiya i antimikrobnaya khimioterapiya [Clinical Microbiology and antimicrobial chemotherapy. 2024;

26(2): 124–140. (In Russ)]. DOI:10.36488/cmac.2024.2.124-140

9. Antimicrobial-resistance. <https://www.who.int/ru/news-room/fact-sheets/detail/antimicrobial-resistance>.

10. AntiMicrobial Resistance Cloud – online platform. <https://amrcloud.net/ru/>

11. Jeong I.S., Song J.Y. Epidemiological Characteristics of Carbapenemase Producing Carbapenem-Resistant Enterobacteriaceae Colonization. Asian Nurs Res (Korean Soc Nurs Sci). 2022; 16(3):134–139. DOI: 10.1016/j.anr.2022.05.002

12. Colín-Castro C.A., et al. The ongoing an-

tibiotic resistance and carbapenemase encoding genotypes surveillance. The first quarter report of the INVIFAR network for 2024. PLoS One. 2025; 20(4). DOI: 10.1371/journal.pone.0319441

13. Van Duin D., et al. Resistant Organism Network Investigators. Molecular and clinical epidemiology of carbapenem-resistant Enterobacteriales in the USA (CRACKLE-2): a prospective cohort study. Lancet Infect Dis. 2020; 20(6):731–741. DOI: 10.1016/S1473-3099(19)30755-8

14. Zaragoza R., Vidal-Cortés P., Aguilar G. Update of the treatment of nosocomial pneumonia in the ICU. Crit Care. 2020; 24(1):383. DOI: 10.1186/s13054-020-03091-2

DOI 10.25789/YMJ.2025.90.06

UDC 575.174

F.M. Teryutin, T.V. Borisova, A.M. Cherdonova, G.P. Romanov, V.G. Pshennikova, A.V. Solovyov, S.A. Fedorova, N.A. Barashkov

ATYPICAL CASES OF HEARING LOSS IN PATIENTS WITH A MITOCHONDRIAL VARIANT m.1555A>G OF THE *MT-RNR1* GENE IN THE REPUBLIC OF BURYATIA

Yakut Science Center of Complex Medical Problems (YSC CMP): **TERYUTIN Fedor Mikhailovich** – PhD, researcher of the Laboratory of Molecular Genetics, e-mail: rest26@mail.ru, ORCID: 0000-0002-8659-0886; **PSHENNIKOVA Vera Gennadievna** – PhD in Biology, visiting research fellow, Laboratory of Molecular Genetics, e-mail: psennikovavera@mail.ru, ORCID: 0000-0001-6866-9462; **BARASHKOV Nikolay Alekseevich** – PhD in Biology, visiting research fellow, head of the Laboratory of Molecular Genetics, e-mail: barashkov2004@mail.ru, ORCID: 0000-0002-6984-7934; M.K. Ammosov North-Eastern Federal University (M.K. Ammosov NEFU): **BORISOVA Tuyara Valeryevna** – junior researcher of the research laboratory of molecular biology, Institute of Natural Sciences, e-mail: borisovaty96@gmail.com ORCID: 0000-0002-5019-067; **CHERDONOVA Alexandra Matveevna** – junior researcher of the research laboratory of molecular biology of IEN e-mail: cherdonovasasha96@gmail.com ORCID: 0000-0002-4168-9516; **ROMANOV Georgiy Prokopyevich** – PhD in Biology, Research Fellow of the Research Laboratory of Molecular Biology, Institute of Natural Sciences, e-mail: gpromanov@gmail.com, ORCID: 0000-0002-2936-5818; **SOLOVYOV Aisen Vasilyevich** – PhD in Biology, Senior Researcher of the Research Laboratory of Molecular Biology, Institute of Natural Sciences, e-mail: nelloann@mail.ru, ORCID: 0000-0003-0914-3609; **FEDOROVA Sardana Arkadievna** – Doctor of Biology, head of the Research Laboratory of Molecular Biology, Institute of Natural Sciences, e-mail: sardanaafedorova@mail.ru, ORCID: 0000-0002-6952-3868.

In a previous study, we found a high prevalence of the m.1555A>G variant of the *MT-RNR1* gene, which causes mitochondrial hearing loss (OMIM 561000) among deaf patients living in the Baikal Lake region. In this regard, in the present study, a genotype-phenotypic analysis of the hearing function in individuals with the m.1555A>G variant was carried out in the discovered Siberian region. Clinical and audiological analysis was performed in 48 people with this mitochondrial variant, whose average age was 51.3±15.5 years. The obtained genotype-phenotypic data are consistent with previously conducted studies of the features of the auditory function in individuals with m.1555A>G, which note incomplete penetrance of the manifestation of the pathological phenotype. Of particular interest in our cohort are three cases of mixed hearing loss, including both sensorineural (inner ear defect) and conductive (middle ear defect) components. The detected conductive component, which is atypical for this mitochondrial form of the disease, may be associated with idiopathic non-infectious foci of the pathological process in the middle ear. We do not exclude the possibility that the detected clinical signs may be a consequence of systemic damage to the hearing organ in this mitochondrial variant. On the other hand, the detected cases may be related to a cross-pathological effect caused by another form of a less common or rare disease. The obtained results require further genotype-phenotypic studies.

Keywords: mitochondrial hearing loss, m.1555A>G variant, *MT-RNR1* gene, genotype-phenotypic analysis, Buryatia

For citation: Teryutin F.M., Borisova T.V., Cherdonova A.M., Romanov G.P., Pshennikova V.G., Solovyov A.V., Fedorova S.A., Barashkov N.A. Atypical cases of hearing loss in patients with a mitochondrial variant m.1555A>G of the *MT-RNR1* gene in the Republic Buryatia. 2025; 90(2): 26–30. <https://doi.org/10.25789/YMJ.2025.90.06>

Introduction. Mitochondria are intracellular organelles responsible for the production of adenosine triphosphate (ATP) through a process called oxidative phosphorylation [20]. In this process, energy is released by breaking down glucose and fatty acids via the mitochondrial respiratory chain [31]. Mutations in mitochondrial DNA have been described primarily in various rare syndromes, but are also found in more common diseases such as sensorineural hearing loss. One such mitochondrial

mutation leading to isolated hearing loss is m.1555A>G in the *MT-RNR1* gene (OMIM 561000). There are several hypotheses regarding the pathogenetic mechanism of m.1555A>G in the *MT-RNR1* gene. In general, researchers believe that the m.1555A>G variant of the *MT-RNR1* gene is one of the “mild” ones compared to other pathogenic variants in mitochondrial DNA, since it does not lead to systemic disorders and does not always lead to hearing loss, and the manifestation of the pathogenic effect of

this mutation requires the participation of modulating factors [12, 22, 24]. Some of these modulators are probably aminoglycoside antibiotics, the principle of action of which is based on their ability to bind to the A-site of the 16S subunit of the bacterial ribosome and thus selectively disrupt the synthesis of prokaryotic proteins without affecting the ribosomes of eukaryotes, due to structural differences [12, 25]. The A>G substitution at position 1555 of human 12S rRNA results in a new C-G pairing, resulting in similarity with the A-site of bacterial 16S rRNA, which is a target for aminoglycoside antibiotics [13]. However, another hypothesis suggests that the m.1555A>G variant of the *MT-RNR1* gene can exhibit a pathogenic effect without the influence of external modulators [10, 12, 13, 16, 23, 25, 30]. Since the substitution of adenine for guanine at position 1555 of the *MT-RNR1* gene results in a change in the conserved A-site (aminoacyl-tRNA acceptor site) of 12S rRNA, this may lead to reading errors during the synthesis of oxidative phosphorylation proteins [23]. In a previous study, we found a high prevalence of the m.1555A>G variant of the *MT-RNR1* gene among patients with hearing impairments living in the Lake Baikal region [15]. With an average worldwide prevalence of the m.1555A>G variant of 1.8% (863/47328), the overall contribution among patients with hearing impairments in the Republic of Buryatia was 12.7% (21/165), and among Buryat patients 20.2% (15/74) [15]. The obtained results indicate that Eastern Siberia is the second largest region of accumulation of the mitochondrial form of hearing loss in the world, after the southern European territory of the Iberian Peninsula, where the overall contribution of this form of hearing loss varies from 17% to 41% [6, 34]. Analysis of the complete mitochondrial genome in 14 unrelated Buryat families carrying the m.1555A>G variant revealed a mitochondrial lineage common to the vast majority of examined individuals, associated with subhaplogroup A5b (92.9%). Considering that more than 90% of Buryat families with the m.1555A>G variant belonged to the same maternal line, it was suggested that the high prevalence of this pathogenic variant in the Lake Baikal region is due to the founder effect [Borisova et al., 2024]. In this regard, in the present work, a genotype-phenotypic analysis of the state of auditory function in patients with the m.1555A>G variant in the *MT-RNR1* gene was carried out in the discovered Siberian focus of accumulation of this mitochondrial disease.

Materials and methods. Study sample. In the Republic of Buryatia, 48 people with the pathogenic variant m.1555A>G of the *MT-RNR1* gene were studied, the average age at the time of the study was 51.3±15.5 years. By nationality, the study sample consisted of: Buryats - 97.9% (47/48), Russians - 2.1% (1/48).

Clinical and audiological analysis. Audiological examination of the hearing state was carried out using pure tone threshold audiometry using an "AA222" audiometer (Interacoustics, Denmark). The degree of hearing loss was assessed by the hearing thresholds of the better hearing ear in the speech frequency range of 0.5, 1.0, 2.0, 4.0 kHz according to the international classification, according to which the first degree of hearing loss corresponds to 26-40 dB, the second degree - 41-55 dB, the third degree - 56-70 dB, the fourth degree - 71-90 dB, deafness >90 dB. For a detailed audiological analysis, we used the clinically important speech frequency range (PTA_{0.5,1,0.2,0.4,0.0kHz}). Audiograms with breaks were normalized by introducing the maximum values (120.0 dB) at frequencies to which the patient did not respond. Sensorineural hearing loss was diagnosed in cases of increased bone and air conduction thresholds on audiograms, mixed - with an increase in bone and air conduction thresholds with an interval exceeding 20.0 dB in total in the PTA_{0.5,1,0.2,0.4,0.0kHz}. Hearing loss was considered asymmetric if the interaural difference in hearing thresholds at the PTA frequencies of 0.5, 1, 0.2, 0.4.0 kHz was more than 15.0 dB.

Detection of the m.1555A>G variant in the *MT-RNR1* gene. Genomic DNA was extracted from venous blood using the phenol-chloroform method. Detection of the m.1555A>G variant in the *MT-RNR1* gene was performed by PCR-RFLP analysis using the previously described sequence of oligonucleotide primers, with a modified reverse primer, which allows the creation of an artificial

recognition site for the restriction endonuclease *HaeIII* [10]. As a result, after 12-hour treatment of the amplification product at 37°C with the *HaeIII* enzyme: normally (1555A) two restriction fragments are formed (216 and 123 bp), with the substitution (1555G) three restriction fragments (216, 93 and 30 bp). Verification of the presence of m.1555A>G in the *MT-RNR1* gene was carried out by Sanger sequencing using the original sequence of oligonucleotide primers: F - AAACGCTTAGCCTAGCCACA, R - GCTACACTCTGGTTCGTCCA, selected using the Primer-BLAST program [29].

Ethical approval. The studies provided for in this work were carried out after informed written consent of the participants. The research work was approved by the local committee on biomedical ethics at the Yakut Scientific Center of Complex Medical Problems in 2019 (Yakutsk, protocol No. 7 dated August 27, 2019).

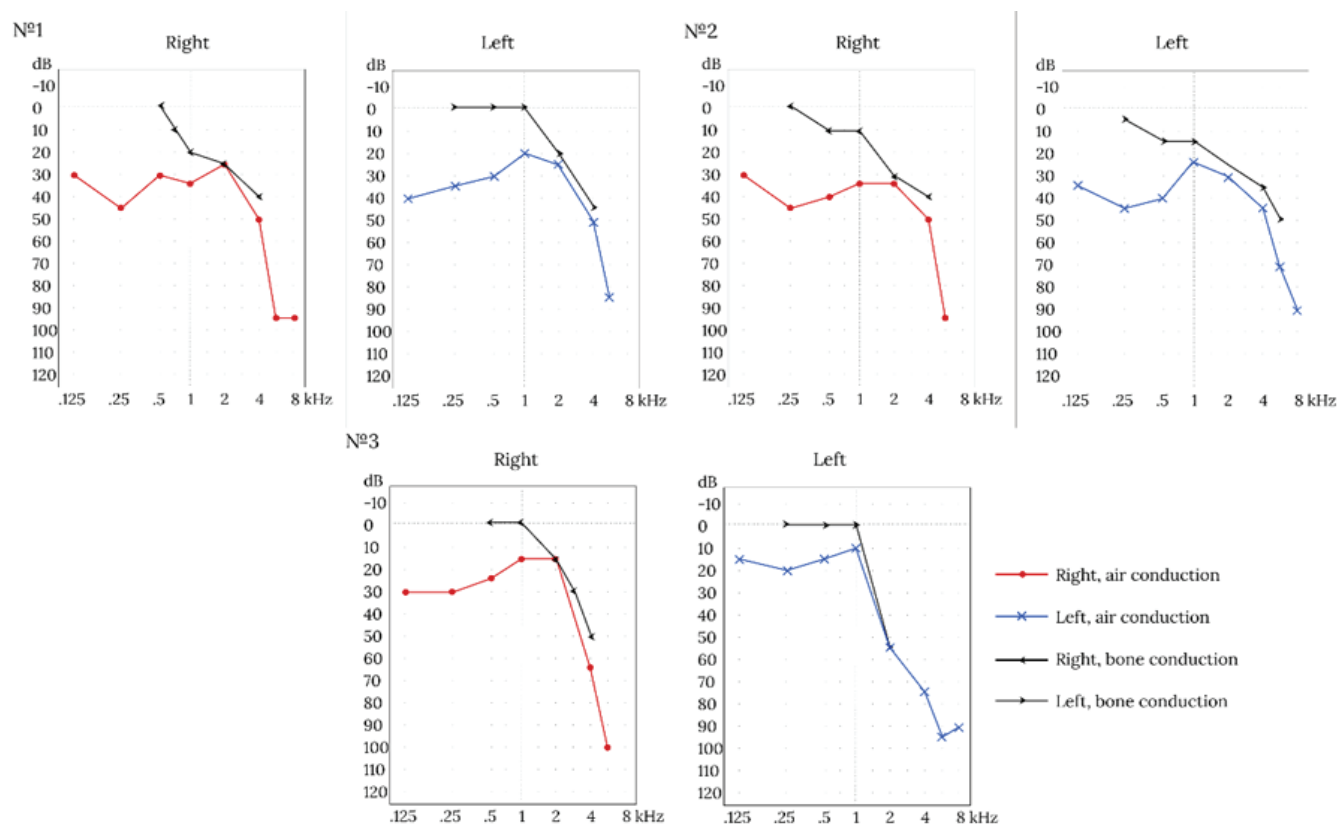
Results and discussion. In this work, a clinical and audiological analysis of the hearing function was performed in 48 individuals with the mitochondrial variant m.1555A>G in the *MT-RNR1* gene (mean age 51.3 years) living in the Republic of Buryatia. As a result, audiological profiles of the state of the hearing function were obtained for all study participants. The characteristics of individuals with m.1555A>G in the *MT-RNR1* gene are presented in Table.

In 27.1% of individuals with the m.1555A>G variant of the *MT-RNR1* gene, hearing in the PTA_{0.5,1,0.2,0.4,0.0kHz} frequency range was within normal values. In 64.6% of the examined subjects, the type of hearing loss was sensorineural, of varying severity from I–II degree of hearing loss to profound (Table). In 8.4% of patients with the m.1555A>G variant of the *MT-RNR1* gene, a mixed form of hearing loss was detected, including both sensorineural (inner ear pathology) and conductive components (middle ear pathology) (Table). One patient with a mixed type of hearing loss was found to have

The auditory function in individuals with m.1555A>G in the *MT-RNR1* gene

Type and degree of the hearing loss	n	%
Normal hearing	13	27.1
Sensorineural hearing loss (n=31. 64.6%)		
- I-II degree	2	4.1
- III-IV degree	9	18.7
- Profound	20	41.6
Mixed hearing loss (n=4. 48.4%)		
- I-II degree	4	8.4
Total	48	100

Note: n - is the number of individuals with m. 1555A>G in the *MT-RNR1* gene.



Audiograms of patients with mixed hearing loss with the mitochondrial variant m.1555A>G in the *MT-RNR1* gene

a perforation of the eardrum, indicating inflammatory processes caused by otitis media. In three of the four patients, signs associated with developmental abnormalities or otitis media were not detected. The age at the time of the examination of these three patients was 54, 68 and 69 years, respectively. All three patients - one man and two women - came from unrelated Buryat families and lived in the same region of the Republic of Buryatia. It should be noted that before the examination, these patients did not complain about their hearing, but according to the clinical and audiological examination, hearing loss in the speech frequency range ($PTA_{0.5,1,0.2,0.4,0.8\text{kHz}}$) corresponded to the first degree of mixed hearing loss. Audiograms of patients with a mixed form of hearing loss with the m.1555A>G variant in the *MT-RNR1* gene are shown in Figure.

The genotype-phenotypic data obtained by us are consistent with previously conducted studies of the characteristics of the hearing function in patients with the m.1555A>G variant in the *MT-RNR1* gene, which note incomplete penetrance of the manifestation of the pathological phenotype in affected families [10, 12, 13, 16, 23, 25, 30, 32]. In other words, not all carriers of m.1555A>G may have clinically significant hearing

loss. It is believed that the age of onset, as well as the degree of hearing loss in individuals with m.1555A>G in the *MT-RNR1* gene, can vary widely - from normal hearing to deafness. The manifestation of signs of hearing loss in carriers of the m.1555A>G variant in some families positively correlated with treatment with aminoglycoside antibiotics [10, 12, 13, 16, 23, 25, 30], as well as with the age of patients [42]. However, not all registered cases of mitochondrial hearing loss can be explained by the ototoxic effect of drugs and progression with age. In this regard, many researchers believe that there are other factors, including genetic

variants in the nuclear genome), modulating the pathological "manifestation" of the m.1555A>G variant [7, 10, 12, 13, 14, 18, 23, 25, 27, 32]. Since we did not find any references in the literature about conductive or mixed type of hearing loss in patients with m.1555A>G in the *MT-RNR1* gene, and the pathogenetic mechanism of mitochondrial hearing loss is associated with damage to cochlear cells, the type of hearing loss should be exclusively sensorineural. In this regard, three cases of mixed type of hearing loss in patients with m.1555A>G in the *MT-RNR1* gene are of particular interest in our cohort of

examined individuals. We do not exclude the possibility that the detected clinical signs may be a consequence of previously undescribed systemic damage to the hearing organ in the mitochondrial variant m.1555A>G. Since, despite the fact that the m.1555A>G variant of the *MT-RNR1* gene is generally considered to be a non-syndromic sensorineural type of hearing loss, there is an alternative opinion that this pathogenic variant is capable of having not only local, but even multi-organ damage potential, which requires a broader clinical study of this form of the disease, since multi-system manifestations can be barely noticeable or even subclinical (short stature, osteopo-

rosis, arterial hypertension and recurrent headache) [11].

On the other hand, our three cases of mixed hearing loss in patients with m.1555A>G in the *MT-RNR1* gene in the Republic of Buryatia may be associated with a cross-over pathological effect caused by another form of a less common or rare disease. Due to the absence of other clinical manifestations in the patients examined by us, as well as their age (54, 68, and 69 years) and gender (one man and two women), it is unlikely that the identified cases are associated with a cross-over effect of an

X-linked recessive form of hearing loss (DFNX2, OMIM 304400). In addition, we cannot completely exclude the version of an atypical manifestation (incomplete penetrance of clinical signs) of one of the rare syndromes in which a mixed type of hearing loss can be observed: branchio-oto-renal syndrome (OMIM 113650), Stickler syndrome (OMIM 108300), Marfan syndrome (OMIM 154700), Treacher Collins syndrome (OMIM 154500), Axenfeld-Rieger syndrome (OMIM 602482) and many others. However, in our opinion, the most likely cause may be an idiopathic non-infectious focus of the pathological process in the middle ear, caused by otosclerosis. The otosclerotic process is based on focal lesions of the bony capsule of the otic labyrinth, with healthy bone being replaced by newly formed porous, spongy, vascular-rich bone tissue during growth, which is why the early stages of otosclerosis are sometimes called otospongiosis. The prevalence of otosclerosis worldwide varies from 1 in 330 in Europe, 1 in 3,300 in Africa, to 1 in 33,000 in Asia [35]. Patients typically have conductive hearing loss, primarily affecting low and mid frequencies, which often progresses to mixed hearing loss. The disease typically manifests in the second, third, or fourth decade and is generally successfully corrected with a combination of surgery and hearing aids [28]. Although both environmental and genetic risk factors have now been identified, the etiology of sporadic cases of otosclerosis remains unknown [5]. An exception are rare familial forms of otosclerosis, segregating according to the autosomal dominant type of inheritance, for some of which genetic loci linked to this disease have been mapped [1, 2, 4, 8, 17, 19, 36, 37]. Despite some success in mapping loci linked to otosclerosis, pathogenic variants in any genes have not been identified to date. However, in 2022, a 15-nucleotide heterozygous deletion in the coding region of the *FOXL1* gene was identified in families with otosclerosis from the Canadian province of Newfoundland and Labrador [3], the product of which is presumably involved in bone remodeling processes in the auditory capsule [26], which was described as causative for the autosomal dominant form of otosclerosis type 11 (OMIM 620576) [3].

Conclusions. Taking into account the atypical for the mitochondrial form of hearing loss - a mixed type of hearing loss in three Buryat patients with m.1555A>G in the *MT-RNR1* gene, which was not associated with otitis media, the detected clinical signs may be a consequence of

a previously undescribed systemic lesion in this mitochondrial variant. On the other hand, the detected clinical signs may be caused by a cross-pathological effect due to another unspecified form of the disease. In our opinion, the most likely cause of the atypical picture of hearing loss in patients with m.1555A>G may be a cross-pathological effect caused by otosclerosis or another rare disease in which a mixed type of hearing loss can be observed (branchio-oto-renal syndrome, Stickler syndrome, Marfan syndrome, Treacher Collins syndrome, Axenfeld-Rieger syndrome and many other syndromes). Given the wide range of nosological forms associated with mixed type of hearing loss, the obtained results require further molecular genetic studies using high-throughput sequencing methods.

The work was carried out within the framework of the research work of the Yakut Scientific Center of Complex Medical Problems "Study of the genetic structure and burden of hereditary pathology in the populations of the Republic of Sakha (Yakutia) and the State assignment of the Ministry of Science and Higher Education of the Russian Federation (FSRG-2023-0003).

The authors declare no conflict of interest in the submitted article.

References

1. Bel Hadj Ali I., Thys M., Beltaief N., et al. A new locus for otosclerosis, OTSC8, maps to the pericentromeric region of chromosome 9. *Hum Genet.* 2008. 123:267–272. <https://doi.org/10.1007/s00439-008-0470-3>
2. Schrauwen I., Weegerink N.J., Fransen E., et al. A new locus for otosclerosis, OTSC10, maps to chromosome 1q41-44. *Clin Genet.* 2011. 79:495–497. <https://doi.org/10.1111/j.1399-0004.2010.01576.x>
3. Abdelfatah N., Mostafa A.A., French C.R., et al. A pathogenic deletion in Forkhead Box L1 (*FOXL1*) identifies the first otosclerosis (OTSC) gene. *Hum Genet.* 2022. 141(3-4):965-979. doi: 10.1007/s00439-021-02381-1.
4. Van Den Bogaert K., Govaerts P.J., Schatteman I., et al. A second gene for otosclerosis, OTSC2, maps to chromosome 7q34-36. *Am J Hum Genet.* 2001. 68:495–500. <https://doi.org/10.1086/318185>
5. Babcock T.A., Liu X.Z. Otosclerosis: From Genetics to Molecular Biology. *Otolaryngol Clin North Am.* 2018. 51:305–318. <https://doi.org/10.1016/j.otc.2017.11.002>
6. Bravo O., Ballana E., Estivill X. Cochlear Alterations in Deaf and Unaffected Subjects Carrying the Deaf-ness-Associated A1555G Mutation in the Mitochondrial 12S rRNA Gene. *Biochem Biophys Res Commun.* 2006. 344, 511–516, doi:10.1016/j.bbrc.2006.03.143.
7. Bykhovskaya Y., Estivill X., Taylor K., et al. Candidate locus for a nuclear modifier gene for maternally inherited deafness. *American journal of human genetics.* 2000. 66(6), 1905–1910. <https://doi.org/10.1086/302914>
8. Brownstein Z., Goldfarb A., Levi H., et al. Chromosomal mapping and phenotypic characterization of hereditary otosclerosis linked to the OTSC4 locus. *Arch Otolaryngol Head Neck Surg.* 2006. 132:416–424. <https://doi.org/10.1001/archotol.132.4.416>
9. Matsunaga T., Kumanomido H., Shiroma M., et al. Deafness due to A1555G mitochondrial mutation without use of aminoglycoside. *Laryngoscope.* 2004. 114. 1085-91. doi: 10.1097/00005537-200406000-00024.
10. Estivill X., Govea G., Barceló E., et al. Familial Progressive Sensorineural Deafness Is Mainly Due to the MtDNA A1555G Mutation and Is Enhanced by Treatment of Aminoglycosides. *Am J Hum Genet.* 1998. 62. 27–35. doi:10.1086/301676.
11. Finsterer J. Variant m.1555A>G in MT-RNR1 causes hearing loss and multiorgan mitochondrial disorder. *Medicine (Baltimore).* 2020 99(6):e18488. doi: 10.1097/MD.00000000000018488.
12. Guan M.X., Fischel-Ghodsian N., Attardi G. Biochemical Evidence for Nuclear Gene Involvement in Phenotype of Non-Syndromic Deafness Associated with Mitochondrial 12S rRNA Mutation. *Hum Mol Genet.* 1996. 5. 963–971. doi:10.1093/hmg/5.7.963.
13. Hamasaki K., Rando R.R. Specific Binding of Aminoglycosides to a Human rRNA Construct Based on a DNA Polymorphism Which Causes Aminoglycoside-Induced Deafness. *Biochemistry.* 1997. 36. 12323–12328, doi:10.1021/bi970962r.
14. Kullar P.J., Gomez-Duran A., Gammage P.A., et al. Heterozygous SSBP1 start loss mutation co-segregates with hearing loss and the m.1555A>G mtDNA variant in a large multigenerational family. *Brain.* 2018. 141(1):55-62. doi: 10.1093/brain/awx295.
15. Borisova T.V., Cherdonova A.M., Pshennikova V.G., et al. High prevalence of m.1555A>G in patients with hearing loss in the Baikal Lake region of Russia as a result of founder effect. *Sci Rep.* 2024. 14(1):15342. doi: 10.1038/s41598-024-66254-z.
16. Rovcanin B., Jancic J., Samardzic J., et al. In Silico Model of mtDNA Mutations Effect on Secondary and 3D Structure of Mitochondrial rRNA and tRNA in Leber's Hereditary Optic Neuropathy. *Exp Eye Res.* 2020. 201. 108277, doi:10.1016/j.exer.2020.108277.
17. Chen W., Campbell C.A., Green G.E., et al. Linkage of otosclerosis to a third locus (OTSC3) on human chromosome 6p21.3-22.3. *J Med Genet.* 2002. 39:473–477. <https://doi.org/10.1136/jmg.39.7.473>
18. Li X., Guan M.X. A human mitochondrial GTP binding protein related to tRNA modification may modulate phenotypic expression of the deafness-associated mitochondrial 12S rRNA mutation. *Mol Cell Biol.* 2002. 22(21):7701-11. doi: 10.1128/MCB.22.21.7701-7711.2002.
19. Tomek M.S., Brown M.R., Mani S.R., et al. Localization of a gene for otosclerosis to chromosome 15q25-q26. *Hum Mol Genet.* 1998. 7:285–290. <https://doi.org/10.1093/hmg/7.2.285>
20. Mitchell P. Coupling of phosphorylation to electron and hydrogen transfer by a chemiosmotic type of mechanism. *Nature.* 1961 Jul 8;191:144-8. doi: 10.1038/191144a0.
21. Fischel-Ghodsian N., Prezant T.R., Chaltrow W.E., et al. Mitochondrial Gene Mutation Is a Significant Predisposing Factor in Aminoglycoside Ototoxicity. *Am J Otolaryngol.* 1997. 18. 173–178. doi:10.1016/s0196-0709(97)90078-8.
22. Fischel-Ghodsian N., Prezant T.R., Bu X., et al. Mitochondrial Ribosomal RNA Gene Mutation in a Patient with Sporadic Aminoglycoside

Ototoxicity. *Am J Otolaryngol.* 1993. 14. 399–403, doi:10.1016/0196-0709(93)90113-I.

23. Hobbie S.N., Bruell C.M., Akshay S., et al. Mitochondrial Deafness Alleles Confer Misreading of the Genetic Code. *Proc Natl Acad Sci USA.* 2008. 105. 3244–3249. doi:10.1073/pnas.0707265105.

24. Prezant T.R., Agopian J.V., Bohlman M.C., et al. Mitochondrial Ribosomal RNA Mutation Associated with Both Antibiotic-Induced and Non-Syndromic Deafness. *Nat Genet.* 1993. 4. 289–294. doi:10.1038/ng0793-289.

25. Hutchin T, Haworth I., Higashi K., et al. Molecular Basis for Human Hypersensitivity to Aminoglycoside Antibiotics. *Nucleic Acids Res.* 1993. 21, 4174–4179, doi:10.1093/nar/21.18.4174

26. Hawkey-Noble A, Pater J.A., Kollipara R., et al. Mutation of foxl1 Results in Reduced Cartilage Markers in a Zebrafish Model of Otosclerosis. *Genes (Basel).* 2022. 13(7):1107. doi:10.3390/genes13071107.

27. Guan M.X., Yan Q., Li X., Bykhovskaya Y., et al. Mutation in TRMU related to transfer RNA modification modulates the phenotypic expres-

sion of the deafness-associated mitochondrial 12S ribosomal RNA mutations. *American journal of human genetics.* 2006. 79(2), 291–302. <https://doi.org/10.1086/506389>

28. Cureoglu S., Schachern P.A., Ferlito A., et al. Otosclerosis: etiopathogenesis and histopathology. *Am J Otolaryngol.* 2006. 27:334–340. <https://doi.org/10.1016/j.amjoto.2005.11.001>

29. Ye J., Coulouris G, Zaretskaya I., et al. Primer-BLAST: A Tool to Design Target-Specific Primers for Polymerase Chain Reaction. *BMC Bioinformatics.* 2012. 13, 134, doi:10.1186/1471-2105-13-134.

30. Greber B.J., Bieri P., Leibundgut M., et al. Ribosome. The Complete Structure of the 55S Mammalian Mitochondrial Ribosome. *Science.* 2015. 348, 303–308, doi:10.1126/science.aaa3872.

31. Rich P.R, Maréchal A. The mitochondrial respiratory chain. *Essays Biochem.* 2010. 47:1–23. doi:10.1042/bse0470001.

32. Gallo-Terán J., Salomón-Felechosa C., González-Aguado R., et al. Sensorineural Hearing Loss in Patients With the m.1555A>G Mutation in the MTRNR1 Gene. *Laryngoscope.* 2024. doi:10.1002/lary.31796.

33. Schuelke M, Krude H, Finckh B, et al. Septo-optic dysplasia associated with a new mitochondrial cytochrome b mutation. *Ann. Neurol.* 2002. 51: 388–392.

34. Bellusci M., Paredes-Fuentes A.J., Ruiz-Pesini E., et al. The Genetic Landscape of Mitochondrial Diseases in Spain: A Nationwide Call. *Genes (Basel).* 2021. 12. 1590. doi:10.3390/genes12101590.

35. Thys M., Van Camp G. Genetics of otosclerosis. *Otol Neurotol.* 2009. 30:1021–1032. <https://doi.org/10.1097/MAO.0b013e3181a86509>

36. Thys M., Van Den Bogaert K., Iliadou V. A seventh locus for otosclerosis, OTSC7, maps to chromosome 6q13-16.1. *Eur J Hum Genet.* 2007. 15:362–368. <https://doi.org/10.1038/sj.ejhg.5201761>

37. Van Den Bogaert K., De Leenheer E.M., Chen W. A fifth locus for otosclerosis, OTSC5, maps to chromosome 3q22-24. *J Med Genet.* 2004. 41:450–453. <https://doi.org/10.1136/jmg.2004.018671>

DIAGNOSTIC AND TREATMENT METHODS

Skryabin E.G.

SACRUM WITH FIVE PAIRS OF SACRAL OPENINGS, AS A PATHOGNOMONIC SYMPTOM OF LUMBOSACRAL TRANSITIONAL VERTEBRAE

DOI 10.25789/YMJ.2025.90.07

UDC 616.711.7

Introduction. Clinical experience shows that during radiological examination of the pelvis, patients are often diagnosed with a sacrum with five pairs of sacral openings, while normally there should be four. Purpose: To which form of dysplasia should cases of diagnosis of the sacrum with five pairs of sacral openings be attributed. Material and methods. The clinical material for the study was the results of computed tomography of the lower lumbar spine and pelvic bones in 78 patients who were diagnosed with a sacrum with five pairs of sacral openings. The CT examination of the patients was carried out on a 128-slice «General Electric» device. Results. The study established that the analyzed group of 78 patients was heterogeneous and consisted of two subgroups. Patients of the first subgroup (52 (66.7%) patients) had fused upper sacral vertebrae by transverse processes to the left and right of the sacral crest. Patients of the second subgroup (26 (33.3%) patients) had similar bone fusion of two upper sacral vertebrae on one side, there was no such concrescence on the contralateral side, and synchondrosis was clearly defined. Discussion. The sacrum with five pairs of sacral openings should be attributed to such a congenital pathology of the lumbosacral junction as lumbosacral vertebrae. It is known that this disease is divided, according to the classification of A.E. Castellvi et al. (1984), into 7 different types. Those clinical observations that were diagnosed in patients of the studied cohort should be attributed to types IIIb (first subgroup, 52 patients) and IV (second subgroup, 26 patients) of the disease. In the clinical picture of each of the types of pathology, vertebrogenic pain syndrome of lumbosacral localization prevails. Conclusion. The presence of such a bright radial symptom of sacral pathology as five pairs of sacral openings allows for timely diagnosis of cases of transitional lumbosacral vertebrae, informing patients about the nature of the disease, and, if necessary, prescribing therapy adequate to the severity of the condition.

Keywords: sacrum, cranial sacral vertebrae, sacral openings, lumbosacral transitional vertebrae.

For citation: Skryabin E.G. Sacrum with five pairs of sacral openings, as a pathognomonic symptom of lumbosacral transitional vertebrae. *Yakut Medical Journal.* 2025; 90(2): 30–32. <https://doi.org/10.25789/YMJ.2025.90.07>

Introduction. In clinical practice, there are often situations when, during

a radiological examination of the pelvis, patients are diagnosed with a sacrum with five pairs of sacral openings [9]. In these cases, the question arises: is this radiological picture normal or is it a variant of pathology [11]. There is no direct answer to this question in modern literary sources [3].

Purpose: To establish which form of dysplasia should be attributed to cases of

diagnosis of a sacrum with five pairs of sacral openings.

Material and methods. The clinical material for the study was the results of computed tomography (CT) of 252 patients aged 12 to 86 years with injuries and diseases of the lower lumbar spine and pelvis, in whom images of the sacrum were "obtained" during the radiological diagnostics. In total, out of 252 patients

SKRYABIN Evgeny Gennadievich – MD, Professor, Department of Traumatology and Orthopedics, Federal State Budgetary Educational Institution of Higher Education, Tyumen State Medical University, ORCID: 0000-0002-4128-6127, skryabineg@mail.ru