

O.E. Dogorova, M.K. Vinokurova, E.S. Pavlova,
V.P. Aleksandrova

THE METHOD OF INDIVIDUALIZED REGIONAL LYMPHOTROPIC THERAPY WITH PHOTOPHORESIS IN PATIENTS WITH MULTIDRUG – RESISTANT PULMONARY TUBERCULOSIS

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We propose a method of individualized regional lymphotropic therapy (RLTT) with photophoresis, as part of multimodality treatment for newly identified destructive multidrug-resistant pulmonary tuberculosis (MDR-TB). The method was developed to meet local specifics of the causative agent, *M. tuberculosis* (MTB), and metabolic characteristics of the indigenous population of the north, which had been elucidated in earlier studies: low level of drug resistance to isoniazid, predominance of slow isoniazid acetylation rate irrespective of administration routes, leading to accumulation of high bactericidal concentrations of isoniazid in blood. The treatment method is suited for patients with low MTB resistance to isoniazid (1 mcg/mL), and slow or medium acetylation rate. Indications and contraindications are specified, as well. The method is protected by patent RU2650633 'Method for treating infiltrative pulmonary tuberculosis with multiple drug resistance' [12]. The proposed method requires administration of anti-tuberculosis drugs standardly recommended for chemotherapy of MDR-TB, market-authorized medical device for laser therapy, and standard equipment for procedure room and laser therapy room. The article describes administration sites, dosages, routes of administration and laser irradiation (photophoresis), drug regimens, and timings for x-ray check-ups. Use of the proposed method resulted in improved treatment efficiency per 6-month period: time to eradication of clinical manifestations shortened by a factor 1.3 (OR 6.31; 95% CI 1.31-30.37); time to culture conversion shortened by a factor of 1.4 (OR 5.1; 95% CI 1.85-14.09); time to cavity healing reduced by a factor of 1.6 (OR 3.42; 95% CI 1.51-7.72); duration of hospitalization needed for intensive phase shortened by a factor 1.2 ($p < 0.01$).

Keywords: tuberculosis, levels of drug resistance to isoniazid, isoniazid inactivation, regional lymphotropic therapy, photophoresis, efficiency.

Introduction. The idea of personalized or individualized approach to patient treatment has been existing since the early days of medicine.

Key to treatment efficiency in tuberculosis (TB) are drug circulation lifetime, and concentrations of medications in blood and in the TB lesions. As a rule, treatment is prescribed based on local bacteriological specifics of the causative agent (MTB), and taking into account metabolic characteristics of the indige-

nous population of the north [2, 5, 11]. According to findings reported by Vinokurova M.K. (2010), multidrug-resistant MTB in the Sakha Republic (Yakutia) showed minor level of resistance to isoniazid (1 mcg/mL) in 92.4% of cases, minor and high levels of resistance to rifampicin (40 and 80 mcg/mL, respectively), while high levels of resistance to isoniazid (10 mcg/mL) and rifampicin (80 mcg/mL) were observed in 7.6% of cases [11].

As is known, one of the biological properties of MTB is its lymphotropism [2, 3, 15]. As far back as the 30s of the 20th century, Shtefko V.G. established bactericidal action of the lymph on the MTB as it enters the lymphatic bed. Later, evidence has been obtained supporting the view, that the lymphatic system of the lungs is not merely the route of infection spread, but also the area of the lungs where healing starts [3, 13]. Prof Y.M. Levin developed the method of regional lymphotropic therapy (RLTT), a method of drug delivery to the lymphatic system, and patients have been treated with it for more than 30 years in all sorts of health facilities. As certain parts of the lymphatic system have been shown to be interconnected with certain organs/tissues, drug administration to lymphatic vessels and nodes at certain, specified body regions enables preservation of optimal drug concentration in the target lesion for 24 hours, allowing achievement of high therapeutic efficacy [4, 9].

Gavriliev S.S. and colleagues have

established that the majority of the population of Yakutia were weak isoniazid acetylators irrespective of administration route, and because of it, accumulated high bactericidal isoniazid concentrations in their blood [5]. Lower therapeutic effect in rapid acetylators, and respectively, higher therapeutic effect in weak acetylators, was observed by number of researchers [16]. The method of photophoresis (low-level laser therapy) has been successfully used to accelerate healing process, as it was shown to have beneficial effect on biochemical reactions, metabolism, microcirculation, and antioxidant activity [10, 6, 7, 8]. Further studies by Gavriliev S.S. and colleagues (2004) demonstrated that intercostal intramuscular administration of isoniazid followed by local photophoresis resulted in increased diffusion of isoniazid in the abnormally altered lesion, and was associated with higher rates of healed destructions in the lungs, and faster culture conversion. Based on the above-mentioned, we developed a method of RLTT for pulmonary MDR-TB, wherein lymphotropic administration of isoniazid with photophoresis is used on top of standard chemotherapy regimen for MDR-TB (regimen IV).

Statistical processing of results was performed using IBM SPSS Statistics 22 software suite. Due to absence of normal distributions for most of the parameters, and due to small study group sizes, measure of central tendency and measure

DOGOROVA Oksana E. - Cand. Sci.(Medicine), TB clinician, O.G. Zakharova Zhigansky Central District Hospital, 12 Oyunsky street, 677027, Zhigansk rural locality, Sakha Republic (Yakutia), Mobile: +7 9681629441, email dogorova2904@mail.ru; **VINOKUROVA Maria K.** - Dr.Sci.(Medicine), Deputy director for organization and science, Research-Practice Center 'Phthisiatry', 93 Petr Alekseev street, 677015, Yakutsk; professor at the Department of Internal Medicine and General Practice, Institute of Medicine, M.K. Ammosov North-East Federal University, Mobile: +7 9644239494, email mkvin61@mail.ru; **PAVLOVA Ekaterina S.** - Cand.Sci.(Medicine), Scientific secretary, Research-Practice Center 'Phthisiatry', 93 Petr Alekseev street, 677015, Yakutsk; Associate professor at the Department of Internal Medicine and General Practice, Institute of Medicine, M.K. Ammosov North-East Federal University, Mobile: +7 9142223530, email esp71@mail.ru, **ALEKSANDROVA Valentina P.** - Patent specialist, Research-Practice Center 'Phthisiatry', 93 Petr Alekseev street, 677015, Yakutsk, Mobile: +7 9243601760.

of spreading are presented as medians (Me) and quartile distributions (Q1; Q3). Comparisons of quantitative variables and qualitative characteristics between groups were done using Kruskal-Wallis test, and Pearson's chi-squared test, respectively.

Material and methods. Drug susceptibility to isoniazid was determined based on urine samples, using absolute concentration method and solid medium, in the presence of 1 mcg/mL and 10 mcg/mL of isoniazid.

Urine samples were tested for isoniazid acetylation using L.I. Grebennik's modification of Wollenberg's test (1961, 1965) in 189 patients. According to procedure description, patients can be referred to one of the following types: slow, intermediate, or rapid inactivators (acetylators). Test result was determined based on amount of active isoniazid excreted in urine. Amounts below 10%, between 11 and 15%, or above 16% were interpreted correspondingly, as rapid, intermediate, or slow acetylators.

Serum isoniazid levels and dynamics were estimated using HPLC (high-performance liquid chromatography) in 96 slow and intermediate acetylators, using different drug administration routes: regional lymphotropic (n=32), intravenous drip (n=28), intramuscular (n=19), and oral (n=17). Blood samples were collected at specified time after drug administration via different routes (regional lymphotropic (1), intravenous drip (2), intramuscular (3), oral (4)): 1.5 hours later (1st sample), 6 hours later (2nd sample), 9 hours later (3rd sample).

Resources required for the proposed treatment include medications for anti-TB chemotherapy, market-authorized medical equipment, and standard equipment for procedure room and for laser therapy room.

1. Laser therapeutic device Uzor-A-2K, 2-channel, supplied with magnet applicator tips (Technical Specification TU TY TBO.290.001; Manufacturer: State-owned enterprise 'Voskhod', Kaluga; Reg. no. 94271-122). Light wavelength: 0.89 μ m; number of channels: 2; pulse power per one channel: 4 W; pulse repetition rate: 80-300 Hz; weight: 5.5 kg; pulse duration: 300 nsec.

2. Medications: 10% isoniazid solution (amp): 5.0 mL; 0.5% novocaine injection solution (amp); heparin injection solution (amp): 5000 U/mL.

Treatment method is protected by patent RU2650633 'Method for treating infiltrative pulmonary tuberculosis with multiple drug resistance' [12].

Results and discussion. Cultures of MTB in all patients exhibited low levels of resistance to isoniazid (1 mcg/mL). As is seen in Table 1, rapid acetylators prevailed mostly among non-indigenous population (43.7%), intermediate acetylators were found at equal proportions among non-indigenous and indigenous population (47.9% and 48.2%), while the

proportion of slow acetylators was reliably larger among indigenous population of Yakutia (45.4%).

As it is seen in Table 2, statistically meaningful differences were apparent in serum isoniazid concentrations after 1.5 hours: in group 1 (regional lymphotropic) compared to groups 3 (intramuscular) and 4 (oral); in group 2 (intravenous drip) versus group 4 (oral). Lowest serum isoniazid concentrations were observed in patients treated via lymphotropic route, and the highest concentrations – in patients who received oral treatment.

After 6 hours, no differences in serum concentrations between groups with different administration routes were noted. Concentrations remained at therapeutic levels irrespective of administration route, but tended to decrease in groups with intravenous drip, intramuscular, and oral administration, compared to lymphotropic route.

After 9 hours, group 4 (lymphotropic route) showed statistically reliable differences in serum isoniazid concentrations, compared to the rest of groups. The sus-

Table 1

Isoniazid acetylation types based on urine tests, among indigenous and non-indigenous population of extreme north

Population	Isoniazid acetylation type						Total	
	Rapid		Intermediate		Slow			
	abc.	%	abc.	%	abc.	%	abc.	%
Indigenous	9	6.4	68	48.2	64	45.4	141	100
Non-indigenous	21	43.7	23	47.9	4	8.4	48	100
Total	30	15.9	91	48.1	68	36.0		
p*	$\chi^2 = 31.87$ df = 1 p = 0.001		$\chi^2 = 0.01$ df = 1 p = 0.894		$\chi^2 = 18.29$ df = 1 p = 0.001		189	100

Note: *p – Pearson's χ^2 test.

Table 2

Serum isoniazid concentrations for different administration routes, mg/L (Me (Q1; Q3))

Collection time	Isoniazid concentration (mg/L) for different administration routes:				p*
	1	2	3	4	
1 st sample	4.2 (3.4; 5.0)	8.0 (5.3; 10.7)	12.5 (9.4; 15.6)	17.1 (23.8; 10.3)	$p_{1-3} < 0.001$ $p_{1-4} < 0.001$ $p_{2-4} = 0.030$
2 nd sample	3.1 (2.0; 4.2)	2.2 (0.9; 3.5)	2.4 (1.0; 3.8)	2.2 (1.0; 3.4)	p = 0.321
3 rd sample	2.2 (1.7; 2.7)	0.8 (0.2; 1.4)	0.8 (0.4; 1.2)	0.8 (0.6; 1.0)	$p_{1-2} = 0.003$ $p_{1-3} = 0.005$ $p_{1-4} = 0.008$
Isoniazid concentration decrease, %					
After 6 hours	27.5 (13.6; 41.4)	74.0 (63.7; 84.3)	79.0 (67.7; 90.3)	78.0 (66.3; 89.7)	$p_{1-2} = 0.012$ $p_{1-3} < 0.001$ $p_{1-4} = 0.002$
After 9 hours	48.5 (39.6; 57.4)	84.3 (74.7; 93.9)	92.6 (88.6; 96.6)	93.7 (89.2; 98.2)	$p_{1-3} < 0.001$ $p_{1-4} < 0.001$

Note: * Kruskal-Wallis test

tained therapeutic serum concentrations were observed in lymphotropic group, compared to decreased concentrations (below 1 mg/L) in groups with intramuscular and oral drug administration.

RLTT with photophoresis is indicated for patients with newly detected extensive destructive pulmonary MDR-TB, with minimally allowed level of resistance to isoniazid equal to 1 mcg/mL, and who are either slow (above 16%) or intermediate (11-15%) isoniazid acetylators.

Contraindications to RLTT include: intolerance/allergy to any agents used in treatment; pyoinflammatory diseases of skin and subcutaneous tissue; pronounced pain syndrome in response to drug administration.

Contraindications to photophoresis include: bleeding or hemoptysis; advanced concurrent diseases or complications (stage IIB-III heart failure, neoplasms, organic injury of central nervous system, stage II-III thyrotoxicosis, diseases of hematopoietic system); pregnancy; diseases associated with pronounced metabolic disturbances (decompensated diabetes mellitus) or dystrophy; disorders of blood.

RLTT with photophoresis was used on top of intensive phase of the chemotherapy regimen for pulmonary MDR-TB (regimen IV). RLTT is performed in combination with isoniazid, a highly lymphotropic anti-TB drug with remarkable capacity to reach and reversibly bind with cell structures of lymphatic tissue. Lymph-stimulating permeator agent is needed to ensure delivery of isoniazid to lymphatic bed. As permeator, we used heparin (1.0 mL, 5000 MU) dissolved in 4.0 mL of 0.5% novocaine solution.

Daily dose (10 mg/kg) of isoniazid solution is administered to subcutaneous tissue, alternating injection sites each time (axillary space; 5th intercostal space near the sternum edge; infraclavicular space in the projection of the joint between the first rib and sternum) 5 times a week, followed by photophoresis, during 25 days.

RLTT is performed in the procedure room, strictly adhering to the standards of asepsis and antiseptics.

Injection sites (axillary space, 5th intercostal space, infraclavicular space) are alternated every day, based on the extent of disease. Also, daily alternation of injection sites helps prevent injection site induration in subcutaneous tissue.

The 1st syringe is filled with 1.0 mL (5000 MU) of heparin and 4.0 mL of 0.5% novocaine solution; the 2nd syringe is filled with daily dose of 10% isoniazid solution (10 mg/kg) and 0.5% novocaine solution till a total amount of 10.0 mL is reached.

Then the injection site is swabbed with alcohol swab, and a subcutaneous injection of heparin is performed, followed by injection of isoniazid solution using the same needle.

Injection to axillary space is performed in sitting position. The patient is sitting on the chair, the arm at the side of injection is placed behind the patient's head. Needle is inserted horizontally, parallel to costal surface, for two thirds of needle's length, stepping 4-5 cm. away from the axillary fossa and 1.5-2 cm. away from the edge of greater pectoral muscle (*musculus pectoralis major*).

Injection to the 5th intercostal space near the edge of the sternum is performed in supine position. The injection site is palpated, then a needle is inserted, stepping 1-1.5 cm. away from the edge of the sternum, subcutaneously, at an acute angle, parallel to the rib, for the half of the needle's length.

Injection to infraclavicular space is performed likewise in supine position, with the patient's head turned the opposite way to the injection side. The joint between the first rib and the manubrium of sternum is palpated, and stepping 1-1.5 cm. away, the needle is inserted subcutaneously, at acute angle, parallel to the rib, for the half of needle's length.

The injection site is covered with alcohol wipe and adhesive strip.

15-20 min. after the injection, body area where isoniazid was injected is further exposed to photophoresis. Photophoresis is performed in laser therapy room (pulse frequency 150 Hz, pulse power 2 W, exposure duration 256 sec.), 5 days a week, during a course of 25 sessions.

Additionally, in the evening (18.30), patients received 0.3 g. of isoniazid, depending on their body weight.

On weekends, and after the completion of the course of RLTT with photophoresis, isoniazid was administered intramuscularly (daily dose of 10 mg/kg) in the morning (10.00), and orally (0.3 mg) in the evening (18.30), depending on patient's body weight.

X-ray and CT checkup was performed 21 days after the completion of RLTT with photophoresis. Based on indications, a course of RLTT with photophoresis was repeated.

The efficiency of the proposed treatment in newly detected infiltrative destructive pulmonary MDR-TB in terms of culture conversion estimated after 6 months of treatment was 89.1% (experiment group) vs. 61.5% (control group) ($p < 0.05$).

Radiological assessment after 6

months of treatment among slow or intermediate acetylators with low levels of resistance to isoniazid (1 mcg/mL) showed reliably higher proportion of healed cavities in group treated with RLTT with photophoresis on top of standard chemotherapy (regimen IV): 74.5% (experiment) vs. 46.2% (control) ($p < 0.05$).

Conclusions.

1. Bacteriological examination of *M. tuberculosis* isolated from patients participating in the study showed that all patients had low levels of resistance to isoniazid (1 mcg/mL).

2. Indigenous residents of Yakutia, in the majority, were either slow (45.4%) or intermediate (48.2%) isoniazid acetylators ($\chi^2 = 18.29$ df = 1 $p = 0.001$).

3. Regional lymphotropic administration of isoniazid in combination with photophoresis allowed more long-lasting bactericidal concentration of isoniazid in serum, assessed 9 hours after drug administration (2.2 (1.7; 2.7) mg/L) vs. baseline (4.2 (3.4; 5.0) mg/L), compared to other administration routes ($p < 0.001$).

4. Lymphotropic administration of isoniazid with photophoresis on top of intensive phase of standard chemotherapy regimen IV, in patients with newly detected infiltrative pulmonary MDR-TB with low resistance to isoniazid, substantially improved treatment efficiency after 6 months of treatment: time to eradication of clinical manifestations shortened by a factor 1.3 (OR 6.31; 95% CI 1.31-30.37); time to culture conversion shortened by a factor of 1.4 (OR 5.1; 95% CI 1.85-14.09); time to cavity healing reduced by a factor of 1.6 (OR 3.42; 95% CI 1.51-7.72); duration of hospitalization needed for intensive phase shortened by a factor 1.2 ($p < 0.01$).

Литература

- Аликова С.К., Ранюк Л.Г., Бурдули Н.М., Тадтаева Д.Я. Гемодинамические типы микроциркуляции и лазерная терапия при хроническом панкреатите в сочетании с метаболическим синдромом. *Терапия*. 2019;(3):60-66. [Alikova S.K., Ranyuk L.G., Burduli N.M., Tadtayeva D.Y. Hemodynamic types of microcirculation and laser therapy in chronic pancreatitis in combination with metabolic syndrome. *Therapy*. 2019;(3):60-66. (In Russ.)]
- Бородин Ю.И. Институт лимфологии и проблемы лимфологии. Бюллетень СО РАМН. 2001;(4):5-11. [Borodin Y.I. Institute of lymphology and problems of lymphology. *Bulletin of SB RAMS*. 2001;(4):5-11. (In Russ.)]
- Винокуров А.С., Соколина И.А., Винокурова О.О. Клинико-рентгенологические особенности лимфогенной диссеминации при туберкулезе легких. *Вестник рентгенологии и радиологии*. 2020;(4):253-262. [Vinokurov A.S., Sokolina I.A., Vinokurova O.O. Clinical and radiographic features of lymphogenous dissemination in pulmonary tuberculosis. *Journal of radiology*

and nuclear medicine. 2020;(4):253-262. (In Russ.)]

4. Выренков Ю.Е., Карандин В.И. Клиническая лимфология. Итоги и перспективы развития. Вестник лимфологии. 2009;(3):25-30. [Vyrenkov Yu.E., Karandin V.I. Clinical lymphology. Summary and prospects for development. *Vestnik limfologii [Bulletin of lymphology]*. 2009;(3):25-30. (In Russ.)]

5. Гаврильев С.С., Винокурова М.К., Илларионова Т.С. Индивидуализированная химиотерапия туберкулеза легких. Якутск; 2003. 128 с. [Gavriilyev S.S., Vinokurova M.K., Illarionova T.S. Individualized chemotherapy of pulmonary tuberculosis. Yakutsk; 2003; 128. (In Russ.)]

6. Гаврильев С.С., Винокурова М.К., Мордовская Л.И. Полупроводниковые лазеры во фтизиатрии. Новые технологии лечения. Новосибирск; 2004. 150 с. [Gavriilyev S.S., Vinokurova M.K., Mordovskaya L.I. Semi-conductor lasers in phthisiatry. *New treatment technologies*. Novosibirsk. 2004; 150 (In Russ.)]

7. Григорьев Ю.Г. Противовирусная электромагнитная и лазерная терапия во фтизиатрии. Туберкулез и социально-значимые заболевания. 2018;(4):68-74. [Grigoriev YU.G. Antifibrotic electromagnetic and laser therapy in phthisiology. *Tuberkulez i social'no znachimye zabolevaniya (Tuberculosis and socially significant diseases)*. 2018;(4):68-74. (In Russ.)]

8. Григорьев Ю.Г., Мишин В.Ю., Григорьев А.Ю. Квантовая терапия при туберкулезе легких. Туберкулез и социально-значимые заболевания. 2019;(1):65-71. [Grigor'ev Yu.G., Mishin V.Yu., Grigor'ev A.Yu. Quantum therapy in pulmonary TB patients. *Tuberkulez i social'no znachimye zabolevaniya (Tuberculosis and socially significant diseases)*. 2019;(1):65-71. (In Russ.)]

9. Губкина М.Ф. Химиотерапия туберкулеза легких у подростков с применением регионального лимфотропного метода лечения : автореф. дис. ... канд. мед. наук. М.; 1996. 20 с. [Gubkina M.F. Chemotherapy of pulmonary

tuberculosis in adolescents using regional lymphotropic treatment method. Extended abstract of Cand.Sci.(Medicine) thesis. Moscow. 1996; 20. (In Russ.)]

10. Малиев Б.М., Егорова И.Л., Сорокина И.А. Лазерные технологии в лечении больных туберкулезом лёгких с сопутствующей патологией. Проблемы туберкулеза. 1998;(3):36-41. [Maliev B.M., Egorova I.L., Sorokina I.A. Laser technologies in treatment of pulmonary tuberculosis with concurrent diseases. *Problemy tuberkuleza*. 1998;(3):36-41. (In Russ.)]

11. Винокурова М.К., и др. Оптимизация организации мониторинга эффективности лечения больных туберкулезом легких с множественной лекарственной устойчивостью в Республике Саха (Якутия): метод. пособие. Якутск: ГУ НПЦ «Фтизиатрия» МЗ РС(Я); 2012. 27 с. [Vinokurova M.K., et al. Optimized management of monitoring of the efficiency of treatment in patients with multidrug-resistant pulmonary tuberculosis in the Sakha Republic (Yakutia): guide. Yakutsk: GU NPTS 'Ftiziatriya' MZ RS(Y) Publ.; 2012. 27 p. (In Russ.)]

12. Догорова О.Е., Винокурова М.К.; патентообладатель Научно-практический центр «Фтизиатрия». Способ лечения инфильтративного туберкулеза легких с множественной лекарственной устойчивостью. Патент № 2650633 Российская Федерация. 16 апреля 2018. [Dogorova O.E., Vinokurova M.K., inventor of the Research-Practice Center 'Phthisiatry', assignee. Method for treating infiltrative pulmonary tuberculosis with multiple drug resistance. Russian Federation patent RU 2650633. 2018 Apr 16. (In Russ.)]

13. Пеленева И.М. Клинико-экспериментальное обоснование оптимизации технологий лимфологического профиля в лечении больных туберкулезом легких: дис. ... д-ра мед. наук. Пермь; 2005. 302 с. [Peleneva I.M. Clinical and experimental foundations for improvement of lymphology-based technologies for treatment of patients with pulmonary tuberculo-

sis. *Dr.Sci.(Medicine) thesis*. Perm; 2005. 302 p. (In Russ.)]

14. Догорова О.Е., и др. Региональная лимфотропная терапия в сочетании с фотофорезом у больных туберкулезом легких с множественной лекарственной устойчивостью: пособие для врачей. Якутск: ГБУ РС(Я) НПЦ «Фтизиатрия» МЗ РС(Я); 2015. 21 с. [Dogorova O.E., et al. Regional lymphotropic therapy in combination with photophoresis in patients with multidrug-resistant pulmonary tuberculosis: guide for physicians. Yakutsk: GBU RS(Y) NPTS 'Ftiziatriya' MZ RS(Y) Publ.; 2015. 21 p. (In Russ.)]

15. Струков А.И., Соловьева И.П. Морфология туберкулеза в современных условиях. М.: Медицина; 1986. 227 с. [Strukov A.I., Solovyova I.P. Morphology of tuberculosis in the contemporary context. Moscow: Meditsina Publ.; 1986. 227 p. (In Russ.)]

16. Хоменко А.Г., Чуканов В.И., Корнеев А.А. Эффективность химиотерапии туберкулеза легких с лекарственно-устойчивыми микобактериями. Проблемы туберкулеза. 1996;(6):42-44. [Khomenko A.G., Chukanov V.I., Korneev A.A. Effectiveness of chemotherapy for drug-resistant pulmonary tuberculosis. *Problemy tuberkuleza (Issues of tuberculosis)*. 1996;(6):42-44. (In Russ.)]

17. Штефко В.Г. Вопросы патологической анатомии лимфогенных фаз туберкулезного процесса легких. Проблемы туберкулеза. 1935;(9):20-40. [Shtefko V.G. Topics in pathologic anatomy of lymphogenic disease phase of pulmonary tuberculosis. *Problemy tuberkuleza (Issues of tuberculosis)*. 1935;(9):20-40. (In Russ.)]

18. Degoma E.M., Rivera G., Lilly S.M., et al. Personalized vascular medicine: individualizing drug therapy. *Vascular Med*. 2011;16(5):391-404. doi: 10.1177/1358863X11422251.

19. Jain K.K. From molecular diagnostics to personalized medicine. *Exp Rev Mol Diagn*. 2002;2(4):299-301. doi: 10.1586/14737159.2.4.299.